



Understanding the aging fly through physiological genetics

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1. Introduction

While multiple factors determine the course of an individual's longevity, increased quality of life, development of health services and absence of war have enabled human populations who profit from these social benefits to live longer. Nonetheless, late-onset deteriorating physical and physiologic changes eventually lead to pathology and death (Balcombe and Sinclair, 2001). To understand and intervene with the negative aspects of the aging process in the human population, biologists investigate the molecular pathways, physiologic changes, genetic regulation, environmental influence and evolutionary origins of aging in a variety of species ranging from unicellular yeast to rodents and mammals (Guarente and Kenyon, 2000).

Energy consumption and energy use is one of the hallmarks of all living organisms (Rolfe and Brown, 1997). Energy is required not only for development and

46 maintenance of a living individual, but also for getting access to food, surviving
47 through environmental stress and reproducing. Given the tight association between
48 such energy exchanges and life, it is not surprising that most of the biology of
49 aging uncovers links with the systems that control energy balance, stress resistance
50 and reproduction (Guarente and Kenyon, 2000). The model organism *Drosophila*
51 *melanogaster* has been instrumental in studies of aging, due to its genetic
52 amenability, its relatively short life span and its wide spread use in the laboratory
53 (Rose, 1999; Helfand and Rogina, 2003). Here, the author tries to integrate recent
54 developments in the field of *Drosophila* aging research into a general conceptual
55 framework for the aging process. The term aging is used to include the phenotypic
56 changes both at early and at later stages of life. These encompass morpho-
57 logical changes in fine structure (Anton-Erxleben et al., 1983; Gartner, 1987)
58 fertility and fecundity (Sgro and Partridge, 1999; Arking et al., 2002), climbing and
59 flying activities (Minois et al., 2001; Marden et al., 2003), learning and memory
60 (Fois et al., 1991; Guo et al., 1996; Savvateeva et al., 1999; Neckameyer et al., 2000).
61 The author proposes that aging of individual multicellular organisms should not
62 be viewed as a mere consequence of cellular aging, even though the latter may
63 influence, necessitate and in some cases determine the aging process. Rather, the
64 existence of systemic regulation, which arises through communication between
65 different organs and is coupled to a genetically determined repertoire of responses,
66 is paramount. In this perspective, physiologic control remains responsive to
67 environmental cues and the extent of genetic determination is shaped during
68 evolution.

70 2. *Drosophila melanogaster* as a model system to study aging: new 71 technical advances

72
73 A large number of laboratories around the world are currently trying to
74 understand the aging process in *Drosophila* (reviews in Rose, 1999; Partridge, 2001;
75 Arking et al., 2002; Tatar et al., 2003) and some aim to extend fly life span, without
76 a concomitant loss of fitness or reproductive potential (Parkes et al., 1998; Kang
77 et al., 2002; Helfand and Inouye, 2002). Longevity is in part under genetic control, as
78 demonstrated by selection experiments (Arking, 1987; Riha and Luckinbill, 1996;
79 Sgro et al., 2000) and variations of longevity and other age-associated traits have
80 been documented in natural populations (Draye et al., 1994; Draye and Lints, 1996).
81 Analysis of the *Drosophila* genome sequence (Adams et al., 2000) is therefore
82 invaluable in current investigations, especially as the existence of fly counterparts
83 for most human disease genes reveal extensive molecular conservation between
84 the two species (Reiter et al., 2001). The technical advancements of site-directed
85 mutagenesis through homologous recombination (Rong and Golic, 2000; Rong et al.,
86 2002) and double stranded RNA silencing (Piccin et al., 2001) enable direct genetic
87 manipulations of candidate aging genes (Bernards and Hariharan, 2001) and are
88 currently being implemented in aging research (Kirby et al., 2002; Egli et al., 2003).
89 In addition, systematic P-element mutagenesis offers defined genetic material to
90 search for candidate genes (Spradling et al., 1999; Peter et al., 2002). In fact two of

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91 the *Drosophila* life-extending genes were isolated through such screens (Lin et al.,
92 1998; Rogina et al., 2000). P-element transposition techniques can also be used to
93 enhance transcription of neighboring genes (Tower, 2000) allowing genetic screening
94 for activities that may extend life span (Seong et al., 2001; Landis et al., 2003) or
95 the direct assessment of specific gene overexpression effects (Orr and Sohal, 1994;
96 Tatar et al., 1997; Parkes et al., 1998; Sun and Tower, 1999; Mockett et al., 1999a,b;
97 Minois et al., 2001; Ruan et al., 2002; Sun et al., 2002).

98 Another important advancement is the use of microarray chip technology (DeRisi
99 et al., 1997) to monitor expression patterns of essentially all *Drosophila* mRNA
100 transcripts during aging (Zou et al., 2000; Pletcher et al., 2002). This approach
101 detected several classes of age-related expression profiles, supporting the earlier
102 proposition that during aging gene expression remains controlled and regulated
103 (Rogina et al., 1998). The observations argue against models of aging that solely
104 implicate stochastic errors in metabolism accumulating with time, because such
105 errors would increase the variability of gene expression at late age (not seen in
106 Pletcher et al., 2002), in the absence of a possibly coupled, but regulated response.
107 Development of a molecular signature for senescence will provide a general marker
108 for the aging process, which can then be used to assess the impacts of treatments
109 or genetic interventions in a definitive manner. Microarray studies also confirm that
110 multiple molecular pathways are coordinately functioning in the aging process.
111 Oxidative stress response, basal metabolism and reproduction-related genes alter
112 their expression during aging in predictable ways. In addition, the immune and
113 general detoxification responses are also regulated in an age-dependent manner,
114 pointing to yet another energy requirement for organisms, namely defense against
115 pathogens.

116 With whole genome analysis expanding in the near future, there is a need for
117 software and databank development to analyze and better present the accumulating
118 data (Hood, 2003). A second challenge is to employ appropriate statistical evaluation
119 of these results (Rose and Long, 2002). As we enter into the genomics era, parallel
120 technical advancements in 2D gel electrophoresis and mass spectrometry make it
121 possible to monitor changes in expression of all proteins as well. However,
122 pioneering use of proteomics in the field of human aging (Toda, 2001; Dierick et al.,
123 2002) has not yet been matched by corresponding work in *Drosophila*.

124 Despite the astonishing evolutionary conservation of many molecular path-
125 ways, findings from *Drosophila* longevity research demand further qualifications
126 before they can be transferred to the human case. *Drosophila* is a poikilotherm (i.e.,
127 changes its body temperature according to the environment), develops in discrete
128 stages, each with different life strategies and metabolism, reproduces by large
129 numbers of progeny, which do not require nurturing, and is post-mitotic in its adult
130 stage. Human physiology is much more elaborate, with a bigger complex brain
131 (Mattson et al., 2002), sophisticated systemic regulation (Schwartz et al., 2000;
132 Baudry et al., 2002) and a significant part of energy use directed towards cultural
133 evolution. However, lessons from *Drosophila* aging remain german in both
134 understanding the evolution of aging and the underlying pathways that control
135 (or are associated) with it.

136 **3. The basic metabolic pathways and their participation in energy allocations**

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Every cellular activity requires energy expenditure. Metabolism is the overall process through which living systems acquire free energy by nutrient oxidation, store it and utilize it. This process consists of interconnected pathways with many intermediate metabolites participating in multiple distinct reactions. Metabolic fates are not randomly determined, but rather depend on regulatory networks, which ensure the presence or absence of different enzymes, their activity or inhibition, their subcellular compartmentalization, as well as the availability of the intermediate moieties (Voet and Voet, 1990). Here, the author briefly discusses the major metabolic pathways, to facilitate examination of the changes that occur after genetic or environmental alterations in energy homeostasis. Such alterations significantly affect aging of the fly, as exemplified by the reverse correlation of external temperature and longevity (Miquel et al., 1976) or by life extension following caloric restriction (Pletcher et al., 2002; Rogina et al., 2002).

The digestive process is not well described in *Drosophila*. Large molecules in food are broken down into simpler units, such as amino acids, sugars, fatty acids, and glycerol. Midgut epithelium absorbs these nutrients and initiates catabolic reactions, in which citric acid cycle metabolites are produced and transported to fat bodies and oenocytes, the principal sites of intermediary metabolism (Rogina et al., 2000; Zinke et al., 2002). The transporter for citric acid cycle intermediates, *Indy* (Rogina et al., 2000; Inoue et al., 2002; Knauf et al., 2002), is expressed in the plasma membrane of these tissues. Intermediate metabolites enter into the citric acid cycle (Fig. 1), the primary pathway of carbohydrate and protein decarboxylation, yielding NADH and FADH₂, which are then used in the mitochondrial electron transport chain to generate ATP (oxidative phosphorylation) (Voet and Voet, 1990). Therefore, continuous flux of the citric acid cycle coupled with oxidative phosphorylation contributes energy in the form of ATP during aerobic metabolism. In between meals, continuous flux of the cycle is ensured by anaplerotic functions (Owen et al., 2002). Glucose provides citric acid cycle intermediates, as it is broken down to pyruvate with concomitant energy release (glycolysis). Amino acids can also be used in the absence of glucose (e.g., during starvation).

In addition to providing reducing power for the generation of ATP, the principal immediate donor of free energy, metabolites of the citric acid cycle are also the biosynthetic precursors of molecules for long-term storage of cellular energy, such as glycogen and fat. Gluconeogenesis is the process by which glucose is formed in a reverse sequence of reactions used in glycolysis. Tissues that are “energy suppliers” (Fig. 1) synthesize and secrete glucose, which is used as an energy source in brain and muscle. Glucose is also used in the pentose phosphate pathway or can be stored as glycogen. The pentose phosphate pathway (pentose shunt) generates NADPH, which is essential for numerous biosynthetic reactions, including synthesis of fatty acids, amino acids and nucleotides. The pentose shunt also generates ribulose-5-phosphate, an essential precursor for DNA and RNA. Finally, citrate (another intermediate of the citric acid cycle) can be converted into fatty acids and stored in the form of lipid triglycerides. Systemic regulation of these reactions is

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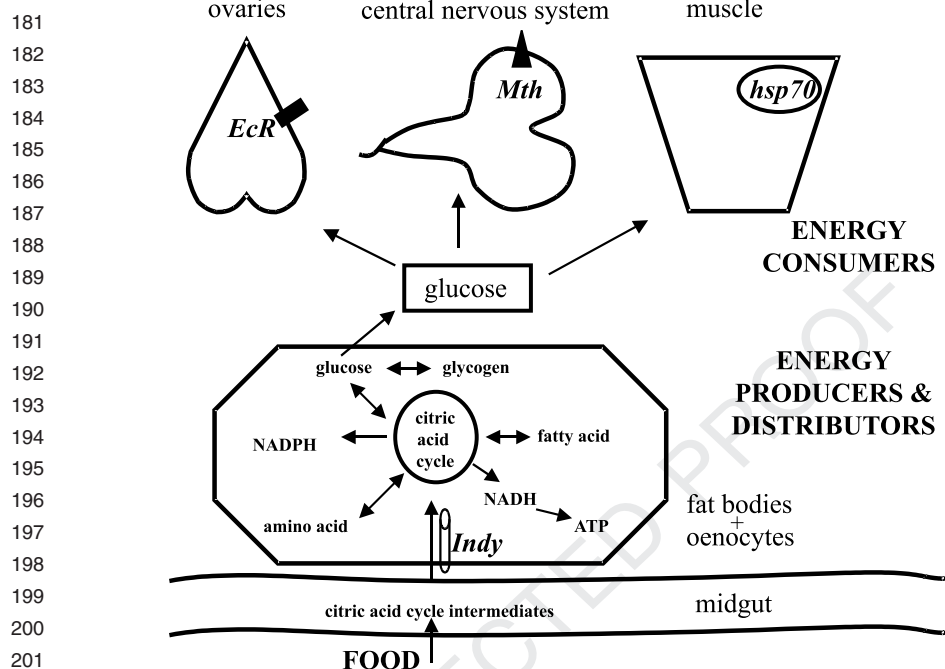


Fig. 1. Midgut, fat bodies and oenocytes are the principal sites of intermediate metabolism. The citric acid cycle allocates energy derived from nutrient oxidation to ATP (for direct use), glucose (to be used in muscle, brain and ovaries), glycogen and fat (storage molecules) and NADPH (reducing power), while generating biosynthetic precursors of the major biomolecules. Regulation of intermediary metabolism is tightly associated with the aging process, as exemplified by extension of life through caloric restriction or genetic interference with the insulin and ecdysone pathways (see text). Candidate “aging” genes are expressed in different tissues. Systemic signaling pathways, in addition to cellular aging, may control the aging process of the adult organism.

controlled by insulin (Baudry et al., 2002; Garofalo, 2002; Rulifson et al., 2002). Thus, the citric acid cycle is at the center of metabolic regulation because of its important roles in generating energy and allocating metabolites towards the different cellular needs.

This constant production of energy involves costs and toxic by-products generated in the electron transport chain of mitochondria (Mandavilli et al., 2002). The superoxide radical is such an example. Superoxide attacks iron sulfur clusters of the very enzymes that perform the citric acid intermediate reactions (Kirby et al., 2002). Effective detoxification rests on the first line antioxidant enzymes superoxide dismutase (Sod) (Phillips et al., 1989), which converts superoxide into hydrogen peroxide, and catalase (Cat) (Griswold et al., 1993), which breaks down hydrogen peroxide into oxygen and water. Hydrogen peroxide is readily diffusible and highly reactive in the presence of iron or copper. Cells employ thiol-dependent antioxidant systems, resting on glutathione and thioredoxins, as a second line of defense against hydrogen peroxide and consequent

226 metal-catalyzed lipid peroxidation (Girotti, 1998; Missirlis et al., 2001). Interestingly,
227 unlike other taxa, insects do not directly depend on glutathione, but rather employ
228 thioredoxin reductase (TrxR) and thioredoxin peroxidases (TPx, Radyuk et al., 2001
229 and GTPx, Missirlis et al., 2003) as their second line of antioxidant defenses (Kanzok
230 et al., 2001). The thioredoxin system requires NADPH to function (Carmel-Harel
231 and Storz, 2000). Thus, part of the energy captured from the pentose shunt is used
232 for self-defense.

233 Oxidative phosphorylation occurs in the highly specialized inner membrane
234 of mitochondria. Citric acid cycle reactions are also predominantly mitochondrial.
235 In contrast, gluconeogenesis and glycolysis, fatty and amino acid metabolism
236 occur in the cytoplasm. Compartmentalization of the different metabolic pathways
237 necessitates the presence of transport systems for the common intermediates in
238 the mitochondrial membrane (Kaplan et al., 1995; Kakhniashvili et al., 1997).
239 Interestingly, many of the citric acid cycle enzymatic activities are present in both
240 the cytosolic and mitochondrial compartments. This raises the intriguing possibility
241 that the enzymes functioning in the mitochondria are geared towards ATP
242 production (NADH and FADH₂), while their cytosolic counterparts may be
243 primarily involved in cataplerotic and anaplerotic functions (Owen et al., 2002).
244 In view of the unanswered questions regarding the involvement of mitochondria in
245 aging (see below), intracellular sites of metabolic regulation will require further
246 clarification.

247 Antioxidant defense enzymes also reside both in the mitochondria and in the
248 cytoplasm of cells. *Drosophila* possesses two *Sod* genes, a cytosolic copper–zinc
249 *Sod1* (Phillips et al., 1989) and a mitochondrial manganese *Sod2* (Duttaroy et al.,
250 1997). *Drosophila* also possess a single *Trxr-1* gene, which encodes two alternative
251 transcripts, one giving rise to a mitochondrial isoform and the other encoding a
252 cytosolic enzyme with identical biochemical properties (Missirlis et al., 2002). Each
253 of the two enzymes provides an essential function; overexpression of TrxR-1 in
254 cytosol cannot compensate for lack of TrxR-1 in mitochondria and vice versa
255 (Missirlis et al., 2002). Thus, in addition to citric acid metabolites, the redox state in
256 cytosol or mitochondria of cells is independently regulated.

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258 4. Oxidative stress and aging

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260 Superoxide and other reactive oxygen species (ROS) lead to DNA damage, lipid
261 peroxidation and protein oxidation (Halliwell and Gutteridge, 1999). Due to these
262 straightforward deleterious effects, ROS are implicated as a causal factor of aging
263 (Harman, 1956; Beckman and Ames, 1998). *Drosophila* has been extensively used to
264 test this theory. Despite recent criticisms (Le Bourg, 2001; Sohal et al., 2002), there
265 is strong supportive evidence for at least some of the theory's proposals. First,
266 mutations in several antioxidant genes greatly impact longevity. This has been
267 demonstrated for cytosolic *Sod1* (Phillips et al., 1989) and mitochondrial *Sod2*
268 (Kirby et al., 2002), cytosolic (Missirlis et al., 2001) and mitochondrial (Missirlis
269 et al., 2002) *Trxr-1*, *catalase* (Griswold et al., 1993) and *glutathione-S-transferase*
270 (Toba and Aigaki, 2000). Furthermore, studies of three marker genes, namely *heat*

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271 *shock protein 70*, *cytochrome oxidase c* and *wingless*, that generally follow an age-
272 specific expression pattern suggest that the actual rate of aging is increased in
273 *Sod1* and *Cat* mutants (Wheeler et al., 1995; Schwarze et al., 1998a; Rogina and
274 Helfand, 2000). A decrease in life span, accompanied by age-related changes in fine
275 structure is also observed if flies are exposed to 50% oxygen levels (Miquel et al.,
276 1975). In summary, ROS overproduction leads to a dramatic acceleration
277 of the aging process.

278 Mitochondria are key sites of ROS generation and energy production (see above)
279 and show distinct age-related morphological changes in flies (Anton-Erxleben et al.,
280 1983). Aged flies show a marked decrease in their ability to produce ATP (Vann and
281 Webster, 1977), but increase hydrogen peroxide production (Ross, 2000) which leads
282 to lipid peroxidation of the membranes (Schwarze et al., 1998b). An interesting
283 linear correlation was noted between mean life span and mitochondrial ROS
284 production between five insect species, including *Drosophila* (Sohal et al., 1995).
285 Mitochondrial DNA remains intact in older flies, but, curiously, mitochondrial
286 transcripts decline sharply with age (Calleja et al., 1993; Schwarze et al., 1998b).
287 What causes this decline remains unknown, but they may involve signaling from the
288 cytoplasm, because cytoplasm derived from old flies and incubated with
289 mitochondria from young individuals has inhibitory effects on ATP production
290 (Vann and Webster, 1977). Thus, dysfunction of mitochondria contributes to cellular
291 and animal aging (Miquel, 1998), though in the case of *Drosophila* not primarily
292 due to DNA mutations.

293 The question that has proven more frustrating to answer conclusively is whether
294 enhanced protection from ROS by overexpression of antioxidant enzymes
295 decelerates aging or not. Some studies confirmed such predictions for both Sods
296 (Parkes et al., 1998; Sun and Tower, 1999; Sun et al., 2002), but in other similar
297 experimental setups life span extension was not observed (Seto et al., 1990; Orr and
298 Sohal, 1993; Mockett et al., 1999a). In addition, overexpression of catalase in an
299 otherwise wild type background did not show positive effects on life span (Orr and
300 Sohal, 1992; Griswold et al., 1993; Sun and Tower 1999; Phillips et al., 2000).
301 Overexpression of a mitochondrially targeted catalase (Mockett et al., 2003) and of
302 thioredoxin reductase (Mockett et al., 1999b) also failed to show any positive effects.
303 In contrast, life span extension was achieved by overexpression of protein repair
304 enzymes, which reduce oxidized methionines (Ruan et al., 2002) and asparagines
305 (Chavous et al., 2001). A correlation of increased antioxidant enzyme activities
306 and longevity was observed in one study of populations that were selected for
307 extended longevities (Arking et al., 2000), but adding to the confusion, inbred lines
308 from the same founding populations exhibited no difference in antioxidant
309 defenses, while retaining their increased longevity, when tested independently
310 (Mockett et al., 2001). These, overall contradicting results may underscore the
311 importance of genetic background differences, tissue specificities, levels of over-
312 expression, subcellular compartmentalization and cofactor requirements of the
313 antioxidant enzyme activities. In other words, negative results from overexpression
314 experiments are not sufficient to discard the otherwise well-documented causal role
315 of ROS in the aging process. Conversely, regarding ROS as the sole contributor to

316 the process is an oversimplification, as many apparently unrelated factors (see below)
317 influence aging as well, and potent detoxification machineries can be employed for
318 protection.

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320 **5. Intermediate metabolism and aging**

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322 Intermediate metabolism allocates energy towards biosynthesis, ATP production
323 or maintenance of an intracellular reduced redox environment (i.e., protection from
324 oxidative stress). Genetic evidence for a tight association between intermediate
325 metabolism and the rate of aging emerges from the analysis of mutations in the *Indy*,
326 *InR* and *chico* genes (Rogina et al., 2000; Clancy et al., 2001; Tatar et al., 2001). *Indy*
327 encodes a membrane transporter for citrate and other dicarboxylic acid cycle
328 intermediates (Inoue et al., 2002; Knauf et al., 2002). Reduced expression of *Indy*,
329 caused by P-element insertions in this locus, results in *Drosophila* strains with
330 extended longevity (Rogina et al., 2000). This observation persists in different genetic
331 backgrounds. Basal metabolic rate as measured by CO₂ emission appears unaltered
332 and no associated decrease in fecundity or locomotion is observed for adequately
333 fed flies (Marden et al., 2003). In contrast, nutritional restriction resulted in a
334 significant downregulation of early fecundity in *Indy* mutants compared to respective
335 wild type controls, demonstrating the importance of environmental variables when
336 evaluating complex phenotypes (see below).

337 *InR* encodes an insulin receptor and *chico* is part of the *InR* signaling cascade
338 (Garofalo, 2002). Reduced expression of either gene also results in extended life span
339 phenotypes (Clancy et al., 2001; Tatar et al., 2001). Insulin signaling is best known
340 for its role in glucose homeostasis (Baudry et al., 2002) and therefore *InR* and
341 *chico* may be part of a control mechanism regulating nutrient availability, which
342 also involves *Indy*. Indeed, caloric restriction extends longevity by an unknown
343 mechanism (Pletcher et al., 2002; Rogina et al., 2002). Microarray analysis from
344 caloric restricted flies suggests a slower progression of the rate of aging, as assessed
345 by a delayed, yet characteristic genome-wide age-dependent expression pattern
346 (Pletcher et al., 2002). Asking if slowing the rate of aging by caloric restriction and
347 by the *chico* mutation occurs by overlapping mechanisms, Clancy et al. reported that
348 *chico* mutants exhibit an optimum life span at a higher food concentration than that
349 of wild type flies (Clancy et al., 2002). The same study concluded that *chico* mutants
350 starve faster and their enhanced longevity respective to wild type strains depends on
351 food availability, arguing for an overlapping mechanism of the genetic and
352 environmental manipulations. Regulation of aging by insulin-like signals extends to
353 other species as well (Tatar et al., 2003).

354 If modulating intermediate metabolism affects aging, the question arises as to
355 whether aging in turn impacts metabolic efficiency. Mitochondrial aconitase, a key
356 iron-sulfur cluster enzyme of the citric acid cycle, is the predominant protein that
357 undergoes oxidative carbonylation with age (Das et al., 2001). This oxidation
358 inactivated the enzyme, an effect also observed in flies with silenced *Sod2*, which have
359 a dramatically reduced life span (Kirby et al., 2002). In addition to these
360 observations, a key step in the electron transport chain mediated by cytochrome c

361 oxidase shows age- and oxidative stress-dependent loss in function (Schwarze et al.,
362 1998a). In summary, both citric acid cycle and oxidative phosphorylation
363 functionally decline in aging *Drosophila*.

364 Finally, classic selection experiments also asserted a tight association between
365 metabolism and longevity (Riha and Luckinbill, 1996; Arking et al., 2002). Analysis
366 of larval metabolism by tracing radioactive glucose incorporated into proteins and
367 lipids shows a directly proportional change in the amount of metabolized food
368 relative to mean life span (Riha and Luckinbill, 1996). In addition, a conspicuous
369 decrease in life spans of previously selected long-lived strains when reared at low
370 population densities can be attributed to greater nutrient intake by those animals.
371 Furthermore, extensive analysis of different longevity phenotypes that have been
372 obtained through various selection regimes has led to the formulation of an
373 integrated interpretation of the changes that eventually lead to extension of life
374 (Arking et al., 2002). The proposed steps include an initial upregulation of
375 antioxidant defenses coupled to an increase in the use of the pentose shunt. This is
376 later followed by alterations in mitochondrial fatty acid composition and other
377 changes necessary to reduce the leakage of hydrogen peroxide from mitochondria
378 into the cytosol. The recaptured energy can be diverted from somatic maintenance
379 back into reproduction. This is an elegant proposal that is consistent with our
380 current understanding of the process and corroborated by many experimental
381 observations mentioned above and below; further investigations are expected to
382 unravel orchestrated pathways that bring about these changes.

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384 6. The interplay between different organs during aging

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386 In multicellular organisms, different cell types are not equally responsive to
387 environmental, genetic or hormonal alterations. Some cells differentiate to perform
388 highly specialized tasks and cannot be replaced if lost or injured. Neurons, for
389 instance, remain highly active, transfer material and electric signals over great
390 distances and sometimes continuously respond to hazardous stimuli, such as UV
391 light. Whether all tissues age at the same time and the extent to which they
392 contribute to senescence and death is not known.

393 In *Drosophila*, genes that when mutated extend life span are preferentially
394 expressed in a variety of different tissues (Fig. 1). Reduced expression levels of a
395 novel G protein-coupled receptor *methuselah* (*mth*) significantly extend fly life span
396 and resistance to stress (Lin et al., 1998). Intriguingly, this receptor is localized at the
397 synapse of fly motorneurons (Song et al., 2002). Human Sod1 overexpression in fly
398 motorneurons is also associated with life span extension, implicating these cells
399 in adult life span control of *Drosophila* (Parkes et al., 1998). *Mth* functions as a
400 positive regulator of pre-synaptic transmission (Song et al., 2002). This provides an
401 elegant mechanism, whereby *mth* could act as a classic antagonistic pleiotropy gene
402 (see next section). In young flies *Mth* upregulates neurotransmitter release,
403 presumably enhancing neuronal responsiveness, which may eventually lead to
404 damage and premature degeneration. This hypothesis predicts that overexpression of
405 *mth* should decrease life span. Song et al. performed an experiment using the *elav*

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406 pan-neuronal driver line to induce *mtH* expression from a *UAS*-transgene and did not
407 observe an impact on fly life span. However, in view of the observation that when the
408 *elav* driver line was used to overexpress human Sod1, a life span increase was not
409 reproduced (Phillips et al., 2000) it seems important to test the flies which
410 overexpress *mtH* via the *D-42* driver line, which expresses specifically in
411 motorneurons. Further investigations of possible genetic interactions between *mtH*
412 and *Sod1* will address if the two genes act in the same process within motorneurons
413 (Parkes et al., 1999).

414 A second single-gene mutant with a dramatic effect on longevity is *Indy* (Rogina
415 et al., 2000). As mentioned above, *Indy* is regulating transport of citric acid cycle
416 intermediates in midgut, fat bodies and oenocytes, therefore acting in organs with
417 very distinct physiological roles from motorneurons (Fig. 1). Another candidate
418 “aging” gene, *heat shock protein 70*, is induced by oxidative stress or aging
419 predominantly in thorax flight muscle (Wheeler et al., 1995), although the role of
420 heat shock proteins in aging still remains controversial (Tatar et al., 1997; King and
421 Tower, 1999; Kurapati et al., 2000; Minois et al., 2001).

422 Flies selected for reproduction at a late age give rise to long-lived populations
423 (Arking, 1987; Sgro et al., 2000). Sterilizing the long-lived strain by X-rays or
424 through a dominant female-sterile genetic mutation will abolish these effects
425 (Sgro and Partridge, 1999). Oxidative stress susceptibility is also associated with
426 increased egg production (Wang et al., 2001), suggesting a trade-off mechanism in
427 energy use. Therefore, reproductive organs may send signals that systemically
428 regulate the individual’s fitness and/or energy use. Interference with systemic
429 signaling pathways can influence longevity, as exemplified by the evolutionarily
430 conserved insulin pathway regulating aging (Tatar et al., 2003). Interestingly, the life
431 span extension of *InR* mutants was abrogated by supplementation of juvenile
432 hormone (Tatar et al., 2001), low levels of which direct the fly into a diapause state
433 (Tatar and Yin, 2001). Moreover, flies with reduced ecdysone synthesis, or ecdysone
434 receptor also live longer, without an apparent deficit in fertility or activity (Simon
435 et al., 2003). Ecdysone is the main steroid hormone acting in flies and could serve as
436 a signal, sent by the gonads, to sustain the organism in good health (Tatar et al.,
437 2003). In support of this proposal, mutations in histone deacetylase *Rpd3*, a
438 downstream target of ecdysone (Tsai et al., 1999), also extend life span (Rogina et al.,
439 2002). In addition, administration of a drug which induces histone acetylation,
440 4-phenylbutyrate, also confers extended longevity (Kang et al., 2002). Microarray
441 analysis of flies treated with 4-phenylbutyrate showed a conspicuous 50-fold increase
442 in *Sod1* levels and moderate increase in other heat shock proteins and antioxidants
443 (Kang et al., 2002). Furthermore, *Sod1* levels were elevated in *InR* and *chico* mutants
444 (Clancy et al., 2001; Tatar et al., 2001). Therefore, we can suggest a model to explain
445 trade-offs between fertility and oxidative stress susceptibility (Wang et al., 2001), in
446 which sterile flies have lower ecdysone levels, reduced expression of *Rpd3* and
447 consequently higher levels of *Sod1*.

448 This model adds another example to the emerging picture that systemic regulation
449 of the aging organism works at a higher organizational level than cellular aging,
450 through the networking of these cells. The fly genome may also contain a program

451 for coordinating energy metabolism, antioxidant defenses and reproduction to
452 support perpetuation of life. As work in the field of developmental biology unraveled
453 a highly complex process of development, similar complexity can be expected during
454 the process of aging.

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7. Evolutionary considerations

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459 The data presented above suggest that coordination of metabolism is tightly
460 associated with the aging process. This may imply that there are genes that will
461 function to increase or decrease the rate of aging. However, current evolutionary
462 theory refutes the existence of genes that evolve to regulate aging and considers aging
463 as a side effect of vital functions (Kirkwood and Austad, 2000; Kirkwood, 2002;
464 Partridge and Gems, 2002). The mutation-accumulation theory (Medawar, 1952)
465 states that the force of counter-selection for deleterious alleles that manifest their
466 phenotypes post-reproductively decreases at that very time and therefore such alleles
467 accumulate in the genome. Thus, after reproductive stages a combination of
468 deleterious genetic mutations contributes to death. The antagonistic pleiotropy
469 theory (Williams, 1957) states that there is a number of genes (or genetic pathways)
470 that can prove beneficial at early stages of life, but the same genes (or the causes of
471 their early actions) may later negatively impact the organism. Such genes would be
472 favored by selection because of the advantages they provide to young individuals
473 seeking to reproduce outweighing the debilitating effects that follow.

474

475 Although the two theories lead to some opposing predictions (Shaw et al., 1999),
476 they are not mutually exclusive, as presence of one type of alleles does not prohibit
477 simultaneous actions of the second type of alleles and both explain aging as a
478 consequence of declining selection pressures after successful reproduction. Evidence
479 in support of both theories is reviewed by Rose (1999) and they are powerful in
480 explaining why in a given species there are limitations to extreme fluctuations in
481 maximum longevity. The two theories are based on the more general theory of
482 natural selection (Darwin, 1859) in its modern synthesis form (Fisher, 1930;
483 Haldane, 1941). Natural selection is undisputedly the strongest force contributing to
484 the evolution of species and genomes within a species. Nonetheless, it requires a pre-
485 existing variation of genetic populations on which it can act. How this variation is
486 achieved in the first place is also an important fundamental process in evolution.
487 Despite his denial of natural selection altogether, Kimura remains the main
488 contributor to the generally accepted notion that radiation and other causes lead to
489 random mutations, which is how variation arises (Kimura, 1968). However,
490 presuming that biological systems are changing randomly neglects the role of
491 environmental effects on evolving variability (besides those explained by natural
492 selection) and the developmental and historical constraints on how genomes (to give
493 one example) may or may not change. A second presumption made by the theories
494 mentioned above, is that natural selection works only at the level of individual
495 organisms, coupled to the premise that it is simultaneously functioning at the level of
496 single genes. This is clearly not sufficient to describe evolution, as selection of species

496 or even ecosystems occurs in nature, for instance in the event of dinosaur extinction,
497 during speciation or during expansion of a desert (Gould, 2002).

498 How are these criticisms of evolutionary theory related to the question *why do*
499 *Drosophila* age? Imagine a scenario in which flies could not age. If they continue to
500 multiply, they would soon consume the resources of their habitat (as happens with
501 bacteria in a culture). For them to survive, they would have to reproduce less. This
502 would result in diminished variability within the species (because reproduction
503 allows recombination and genetic shuffling). It would increase the chances of being
504 eaten by a bird before reproducing. It would also limit the energy resources of the
505 species, as larvae feed within a fruit and have a metabolism geared towards growth
506 in contrast to the adult imago which searches an appropriate partner and habitat
507 to deposit its eggs. Aging may have evolved exactly to tune such problems of
508 ecological balance and may in this respect be a programmed event, subject to
509 selection, rather than solely a by-product of other functions. In such a context
510 phenomena greatly influencing aging such as diapause (Tatar and Yin, 2001), can be
511 viewed as switches of the metabolic program that will or will not lead to senescence
512 (Kenyon, 2001).

513 The notion that there is a need to transcend (but not refute) current theory was
514 recently proposed by scientists actively working on this field (Promislow and
515 Pletcher, 2002). These authors present other evolutionary parameters that need to be
516 incorporated in theoretical models, such as conflict of sexes or social behavior and
517 discuss the corresponding progress in mathematical modeling. Evolutionary
518 debates set aside, there is a general consensus that organisms have to deal with
519 costs when using their energy sources for their metabolic maintenance and
520 reproduction, and failure to do so is the universal trademark of senescence. Which
521 genetic factors determine different life expectancies of the various species remains
522 at this point obscure. As a more thorough investigation of the biology of aging
523 proceeds and efforts to postpone its manifestations of senescence succeed, we will
524 hopefully gain insight on their origin and purpose.

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526

527 8. Conclusions

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529 Aging experiments in *Drosophila* have shown:

530

- 531 1. ROS are a major causal factor of the aging process.
- 532 2. Decreased metabolic rate (as a result of caloric restriction, temperature drop or
533 genetic mutation) extends life span.
- 534 3. Suppression of fertility extends life span.
- 535 4. Extension of life span is commonly accompanied with corresponding trade-offs in
536 fecundity or metabolic activity.
- 537 5. Selection experiments can result in incremental physiologic changes (observed at
538 different generations) which are heritable and extend life span without apparent
539 trade-offs in fecundity or metabolic activity.
- 540 6. Genetic manipulations of single genes can result in extended life span without
apparent trade-offs in fecundity or metabolic activity.

541 7. Aging is in part under neuroendocrine and hormonal control and gene expression
542 remains regulated even in old animals.

543 These conclusions imply that the aging process is much more complex and
544 interconnected with other physiological functions, than previously anticipated. The
545 roles of different tissues and cellular pathways that are directly affecting the aging
546 individual are just starting to be recognized.
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